

Package ‘CpGassoc’

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Type Package

Title Association Between Methylation and a Phenotype of Interest

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Author Barfield, R., Conneely, K., Kilaru, V

Maintainer R Barfield <barfieldrichard8@gmail.com>

Description

Is designed to test for association between methylation at CpG sites across the genome and a phenotype of interest, adjusting for any relevant covariates. The package can perform standard analyses of large datasets very quickly with no need to impute the data. It can also handle mixed effects models with chip or batch entering the model as a random intercept. Also includes tools to apply quality control filters, perform permutation tests, and create QQ plots, manhattan plots, and scatterplots for individual CpG sites.

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CpGassoc-package	<i>Association Between Methylation and a Phenotype of Interest</i>
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Description

Is designed to test for association between methylation at CpG sites across the genome and a phenotype of interest, adjusting for any relevant covariates. The package can perform standard analyses of large datasets very quickly with no need to impute the data. It can also handle mixed effects models with chip or batch entering the model as a random intercept. Also includes tools to apply quality control filters, perform permutation tests, and create QQ plots, manhattan plots, and scatterplots for individual CpG sites.

There is a tutorial for CpGassoc available at the website [<genetics.emory.edu/conneely/>](http://genetics.emory.edu/conneely/)

Details

Package: CpGassoc Type: Package Title: Association between Methylation and a phenotype of interest Version: 2.60 Date: 20

CpGassoc is a suite of R functions designed to perform flexible analyses of methylation array data. The two main functions are `cpg.assoc` and `cpg.perm`. `cpg.assoc` will perform an association test between the CpG sites and the phenotype of interest. Covariates can be added to the model, and can be continuous or categorical in nature. `cpg.assoc` allows users to set their own false discovery rate threshold, to transform the beta values to $\log(\beta/(1-\beta))$, and to subset if required. `cpg.assoc` can also fit a linear mixed effects model with a single random effect to control for possible technical difference due to batch or chip. `cpg.assoc` uses the Holm method to determine significance. The user can also specify an FDR method to determine significance based on the function `p.adjust` or `qvalue`. `cpg.perm` performs the same tasks as `cpg.assoc` followed by a permutation test on the data, repeating the analysis multiple times after randomly permuting the main phenotype of interest. The user can specify the seed and the number of permutations. If over one hundred permutations are performed QQ plots can be created with empirical confidence intervals based on the permuted t-statistics. For more information see [plot.cpg.perm](#). For more information on how to perform `cpg.assoc` or `cpg.perm` see their corresponding help pages. CpGassoc can also perform quality control (see [cpg.qc](#)).

Author(s)

Barfield, R.; Kilaru,V.; Conneely, K.
 Maintainer: R. Barfield: [<rbarfield01@fas.harvard.edu>](mailto:rbarfield01@fas.harvard.edu)

See Also

[cpg.assoc](#) [cpg.combine](#) [cpg.perm](#) [cpg.work](#) [plot.cpg](#) [scatterplot](#) [manhattan](#) [plot.cpg.perm](#)
[cpg.qc](#)

Examples

```
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true
###This will involve partitioning up the data and performing more gc() to clear up space
###Using cpg.assoc:
data(samplecpg,samplepheno,package="CpGassoc")
results<-cpg.assoc(samplecpg,samplepheno$weight,large.data=FALSE)
results

##Using cpg.perm:
Testperm<-cpg.perm(samplecpg[1:200,],samplepheno$weight,data.frame(samplepheno$Dose),
  seed=2314,nperm=10,large.data=FALSE)
Testperm

#For more examples go to those two pages main help pages.
```

Class <i>cpg.perm</i>	<i>Methods for object of class "cpg.perm".</i>
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Description

Methods and extra functions for class "cpg.perm". `plot.cpg.perm` creates a QQ plot based on the association p-values or t-statistics from the function `cpg.perm`.

Usage

```
## S3 method for class 'cpg.perm'
plot(x, save.plot = NULL, file.type = "pdf", popup.pdf = FALSE,
  main.title = NULL, eps.size = c(5, 5), tplot = FALSE, perm.ci = TRUE, classic = TRUE,
  gc.p.val = FALSE, gcdisplay = FALSE, ...)

## S3 method for class 'cpg.perm'
summary(object,...)

## S3 method for class 'cpg.perm'
print(x,...)

## S3 method for class 'cpg.perm'
sort(x,decreasing,...)
```

Arguments

<code>x</code>	Output from <code>cpg.perm</code> . Of class " <code>cpg.perm</code> ".
<code>save.plot</code>	Name of the file for the plot to be saved to. If not specified, plot will not be saved.
<code>file.type</code>	Type of file to be saved. Can either be " <code>pdf</code> " or " <code>eps</code> ". Selecting <code>file.type="eps"</code> will result in publication quality editable postscript files that can be opened by Adobe Illustrator or Photoshop.
<code>popup.pdf</code>	TRUE or FALSE. If creating a pdf file, this indicates if the plot should appear in a popup window as well. If running in a cluster-like environment, best to leave FALSE.
<code>main.title</code>	Main title to be put on the graph. If NULL one based on the analysis will be used
<code>eps.size</code>	Vector indicating the size of .eps file (if creating one). Corresponds to the options <code>horizontal</code> and <code>height</code> in the <code>postscript</code> function.
<code>tplot</code>	Logical. If TRUE, ordered t-statistics will be plotted against their expected quantities. If FALSE (default), $-\log(p)$ will be plotted. If <code>indep</code> is a class variable this option will be ignored.
<code>perm.ci</code>	Logical. If TRUE, the confidence intervals computed will be from the permuted values, otherwise will be based on the theoretical values.
<code>classic</code>	Logical. If TRUE, a classic qq-plot will be generated, with all p-values plotted against predicted values (including significant). If FALSE Holm-significant CpG sites will not be used to compute expected quantiles and will be plotted separately.
<code>gc.p.val</code>	Logical. If true, plot will use the genomic control adjusted p-values.
<code>gcdisplay</code>	Logical. If true, plot will display the genomic control value in the legend.
<code>object</code>	Output of class " <code>cpg.perm</code> " from " <code>cpg.perm</code> ".
<code>decreasing</code>	logical. Should the sort be increasing or decreasing? Not available for partial sorting.
<code>...</code>	Arguments to be passed to methods, such as graphical parameters.

Note

Empirical confidence intervals will be computed only if there are a hundred or more permutations. Otherwise the theoretical confidence intervals will be plotted.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.perm](#) [plot.cpg](#) [scatterplot](#) [manhattan](#) [cpg.assoc](#)

Examples

```

data(samplecpg,samplepheno,package="CpGassoc")
##We will do the analysis on a subset to save time
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
#The qq plot:
Testperm<-cpg.perm(samplecpg[1:300,],samplepheno$weight,seed=2314,nperm=10,large.data=FALSE)
plot(Testperm)
#The t-statistic plot from cpg.perm has confidence intervals since we were allowed
#to perform permutations on the T-values.
plot(Testperm,tplot=TRUE)
#If there was 100 or more permutations, there would be emperical confidence intervals.

#Getting an example of the non classic QQ plot
plot(Testperm,classic=FALSE)

###Now for Sort
head(sort(Testperm)$results)
head(Testperm$results)

```

cpg

Methods for object of class "cpg"

Description

Methods and extra functions for class "cpg". `plot.cpg` creates a QQ plot based on the association p-values or t-statistics from the function `cpg.assoc`.

Usage

```

## S3 method for class 'cpg'
plot(x, save.plot = NULL, file.type = "pdf", popup.pdf = FALSE,
      tplot = FALSE, classic = TRUE,main.title = NULL, eps.size = c(5, 5),
      gc.p.val = FALSE, gcdisplay = FALSE, ...)

## S3 method for class 'cpg'
summary(object,...)

## S3 method for class 'cpg'
print(x,...)

## S3 method for class 'cpg'
sort(x,decreasing,...)

```

Arguments

<code>x</code>	Output of class "cpg" from <code>cpg.assoc</code> or <code>cpg.work</code> .
<code>save.plot</code>	Name of the file for the plot to be saved to. If not specified, plot will not be saved.
<code>file.type</code>	Type of file to be saved. Can either be "pdf" or "eps". Selecting <code>file.type="eps"</code> will result in publication quality editable postscript files that can be opened by Adobe Illustrator or Photoshop.
<code>popup.pdf</code>	TRUE or FALSE. If creating a pdf file, this indicates if the plot should appear in a popup window as well. If running in a cluster-like environment, best to leave FALSE.
<code>tplot</code>	Logical. If TRUE, t-statistics will be plotted vs. their expected quantiles. If FALSE (default), $-\log(p)$ will be plotted. (Note: if <code>class(x\$indep)='factor'</code> this option will be ignored.)
<code>classic</code>	Logical. If TRUE, a classic qq-plot will be generated, with all p-values plotted against predicted values (including significant). If FALSE Holm-significant CpG sites will not be used to compute expected quantiles and will be plotted separately.
<code>main.title</code>	Main title to be put on the graph. If NULL one based on the analysis will be used.
<code>eps.size</code>	Vector indicating the size of .eps file (if creating one). Corresponds to the options <code>horizontal</code> and <code>height</code> in the <code>postscript</code> function.
<code>gc.p.val</code>	Logical. If true, plot will use the genomic control adjusted p-values.
<code>gcdisplay</code>	Logical. If true, plot will display the genomic control value in the legend.
<code>object</code>	Output of class "cpg" from <code>cpg.assoc</code> or <code>cpg.work</code> .
<code>decreasing</code>	logical. Should the sort be increasing or decreasing? Not available for partial sorting.
<code>...</code>	Arguments to be passed to methods, such as graphical parameters.

Value

`sort.cpg` returns an item of class "cpg" that is sorted by p-value. `summary.cpg` creates a qq-plot based on the data, and scatterplots or boxplots for the top sites.

Note

Plots with empirical confidence intervals based on permutation tests can be obtained from `cpg.perm`. See [plot.cpg.perm](#) for more info.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.perm](#) [cpg.assoc](#) [scatterplot](#) [manhattan](#) [plot.cpg.perm](#)

Examples

```
##Using the results from the example given in cpg.assoc.
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
##QQ Plot:
data(samplecpg,samplepheno,package="CpGassoc")
test<-cpg.assoc(samplecpg,samplepheno$weight,large.data=FALSE)
plot(test)
##t-statistic plot:
plot(test,tplot=TRUE)

#Getting our plot:
plot(test,classic=FALSE)

##Now an example of sort
head(sort(test)$results)

##Summary
summary(test)
```

cpg.assoc

Association Analysis Between Methylation Beta Values and Phenotype of Interest

Description

Association Analysis Between Methylation Beta Values and Phenotype of Interest. For more detail see Tutorial.

Usage

```
cpg.assoc(beta.val, indep, covariates = NULL, data = NULL, logit.transform = FALSE,
chip.id = NULL, subset = NULL, random = FALSE, fdr.cutoff = 0.05, large.data = TRUE,
fdr.method = "BH", logitperm = FALSE)
```

Arguments

beta.val	A vector, matrix, or data frame containing the beta values of interest (1 row per CpG site, 1 column per individual).
indep	A vector containing the variable to be tested for association. cpg.assoc will evaluate the association between the beta values (dependent variable) and indep (independent variable).
covariates	A data frame consisting of additional covariates to be included in the model. covariates can also be specified as a matrix if it takes the form of a model matrix with no intercept column, or can be specified as a vector if there is only one covariate of interest. Can also be a formula(e.g. ~cov1+cov2).

<code>data</code>	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in <code>data</code> , the variables are taken from the environment from which <code>cpg.assoc</code> is called.
<code>logit.transform</code>	logical. If TRUE, the logit transform of the beta values $\log(\text{beta.val}/(1-\text{beta.val}))$ will be used. Any values equal to zero or one will be set to the next smallest or next largest value respectively; values <0 or >1 will be set to NA.
<code>chip.id</code>	An optional vector containing chip, batch identities, or other categorical factor of interest to the researcher. If specified, chip id will be included as a factor in the model.
<code>subset</code>	An optional logical vector specifying a subset of observations to be used in the fitting process.
<code>random</code>	Logical. If TRUE, 'chip.id' will be included in the model as a random effect, and a random intercept model will be fitted. If FALSE, 'chip.id' will be included in the model as an ordinary categorical covariate, for a much faster analysis.
<code>fdr.cutoff</code>	The desired FDR threshold. The default setting is .05. The set of CpG sites with $\text{FDR} < \text{'fdr.cutoff'}$ will be labeled as significant.
<code>large.data</code>	Logical. Enables analyses of large datasets. When <code>large.data=TRUE</code> , <code>cpg.assoc</code> avoids memory problems by performing the analysis in chunks.
<code>fdr.method</code>	Character. Method used to calculate False Discovery Rate. Choices include any of the methods available in <code>p.adjust()</code> or "qvalue" for John Storey's qvalue method (requires that qvalue package is installed). The default method is "BH" for the Benjamini & Hochberg method.
<code>logitperm</code>	Logical. For internal use only.

Details

`cpg.assoc` is designed to test for association between an independent variable and methylation at a number of CpG sites, with the option to include additional covariates and factors.

`cpg.assoc` assesses significance with the Holm (step-down Bonferroni) and FDR methods.

If `class(indep)='factor'`, `cpg.assoc` will perform an ANOVA test of the variable conditional on the covariates specified. Covariates, if entered, should be in the form of a data frame, matrix, or vector. For example, `covariates=data.frame(weight,age,factor(city))`. The data frame can also be specified prior to calling `cpg.assoc`. The covariates should either be vectors or columns of a matrix or data.frame.

`cpg.assoc` is also designed to deal with large data sets. Setting `large.data=TRUE` will make `cpg.assoc` split up the data to enable efficient analysis of large datasets.

Value

`cpg.assoc` will return an object of class "cpg". The functions `summary` and `plot` can be called to get a summary of results and to create QQ plots.

`results` A data frame consisting of the t or F statistics and P-values for each CpG site, as well as indicators of Holm and FDR significance. CpG sites will be in the same

	order as the original input, but the <code>sort()</code> function can be used directly on the <code>cpg.assoc</code> object to sort CpG sites by p-value.
<code>Holm.sig</code>	A list of sites that met criteria for Holm significance.
<code>FDR.sig</code>	A data.frame of the CpG sites that were significant by the FDR method specified.
<code>info</code>	A data frame consisting of the minimum P-value observed, the FDR method that was used, the phenotype of interest, the number of covariates in the model, the name of the matrix or data frame the methylation beta values were taken from, the FDR cutoff value and whether a mixed effects analysis was performed.
<code>indep</code>	The independent variable that was tested for association.
<code>covariates</code>	Data.frame or matrix of covariates, if specified (otherwise NULL).
<code>chip</code>	chip.id vector, if specified (otherwise NULL).
<code>coefficients</code>	A data frame consisting of the degrees of freedom, and if object is continuous the intercept effect adjusted for possible covariates in the model, the estimated effect size, and the standard error. The degrees of freedom is used in <code>plot.cpg</code> to compute the genomic inflation factors.

Author(s)

Barfield, R.; Conneely, K.; Kilaru, V.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.work](#) [cpg.perm](#) [plot.cpg](#) [scatterplot.cpg](#) [combine](#) [manhattan](#) [plot.cpg.perm](#) [sort.cpg.perm](#)
[sort.cpg](#) [cpg.qc](#) [cpg.GC](#)

Examples

```
# Sample output from CpGassoc
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
data(samplecpg, samplepheno, package="CpGassoc")
results<-cpg.assoc(samplecpg, samplepheno$weight, large.data=FALSE)
results
#Analysis with covariates. There are multiple ways to do this. One can define the
#dataframe prior or do it in the function call.
test<-cpg.assoc(samplecpg, samplepheno$weight, data.frame(samplepheno$Distance,
samplepheno$Dose), large.data=FALSE)
# or
covar<-data.frame(samplepheno$Distance, samplepheno$Dose)
test2<-cpg.assoc(samplecpg, samplepheno$weight, covar, large.data=FALSE)

#Doing a mixed effects model. This does take more time, so we will do a subset of
#the samplecpg
randtest<-cpg.assoc(samplecpg[1:10,], samplepheno$weight, chip.id=samplepheno$chip,
random=TRUE, large.data=FALSE)
```

cpg.combine

Combine various objects of class "cpg"

Description

Takes a list containing objects of class "cpg" and combines them into one cpg item. Assumes that there are no repeated CpG sites between the various objects (i.e. analysis wasn't performed on the same sites twice).

Usage

```
cpg.combine(allvalues, fdr.method="BH", fdr.cutoff=.05)
```

Arguments

allvalues	A list containing the "cpg" objects that are desired to be consolidated.
fdr.method	FDR method that user wants to use. For options see the cpg.assoc help page.
fdr.cutoff	The desired FDR threshold. The default setting is .05. The set of CpG sites with FDR < fdr.cutoff will be labeled as significant.

Value

info.data	An object of class "cpg" that is the consolidated version of the objects of class cpg that were passed in.
-----------	--

Note

This is designed to be used by cpg.assoc when it does analysis on large data sets or by the user if they split up the analysis by chromosome or some other such partition.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.assoc](#) [cpg.perm](#) [cpg.work](#) [plot.cpg](#) [scatterplot](#) [manhattan](#) [plot.cpg.perm](#)

Examples

```
data(samplecpg, samplepheno, package="CpGassoc")
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
test1<-cpg.assoc(samplecpg[1:100,], samplepheno$weight, large.data=FALSE)
test2<-cpg.assoc(samplecpg[101:200,], samplepheno$weight, large.data=FALSE)
bigtest<-list(test1, test2)
```

```
overall<-cpg.combine(bigtest)
overall
```

cpg.everything *Multi-Task function*

Description

A function designed to do a group of smaller functions required for the `cpg.assoc`

Usage

```
cpg.everything(x, ...)
```

Arguments

x	Just a generic object
...	Arguments to be passed to methods.

Details

A function created to do a bunch of much smaller tasks within **CpGassoc** based on the class of x.

Note

`cpg.everything` is designed to perform a multitude of smaller tasks for `cpg.assoc` that do not warrant a full help page. These include: warnings, getting the name of the independent variable, designing the random function to be used, and getting the names for the values returned.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.assoc](#) [cpg.perm](#) [plot.cpg](#) [scatterplot](#) [manhattan](#) [plot.cpg.perm](#)

Examples

```
#Has four methods: character, complex, numeric/matrix, and logical
#They correspon to getting the indep variable name, warnings, getting the random function,
#and getting the names for the values returned. For the design of these functions see
#the R code.
```

cpg.GC

cpg.GC and methods for output of function

Description

cpg.GC accepts an object of class "cpg.perm" or "cpg" and returns information regarding Holm and FDR-significance of the GC (genomic control) adjusted test statistics. For "cpg.perm" will return permutation p-values based on the GC-adjusted values from each permutation.

Usage

```
cpg.GC(x)

## S3 method for class 'cpg.gc'
print(x,...)

## S3 method for class 'cpg.perm.gc'
print(x,...)
```

Arguments

x Object of class "cpg.perm" or "cpg".
... Arguments to be passed to methods, such as graphical parameters.

Details

cpg.GC will display the number of Holm and FDR-significant sites using the genomic control adjusted p-values test statistics. It will also display the estimated genomic control inflation factor.

Value

cpg.GC returns an object of class "cpg.gc" or "cpg.perm.gc"

gc.results Matrix consisting of GC-adjusted test statistics for each CpG site. Similar to the results output of cpg.assoc.

gc.info Data frame with information on the number of Holm and FDR significant sites. Will also have the genomic control inflation estimate. Objects from "cpg.perm" will also have information concerning the permutation p-values.

Author(s)

Barfield, R.; Kilaru,V.; Conneely, K.
Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.work](#) [cpg.permplot](#) [cpg.scatterplot](#) [cpg.combine](#) [manhattanplot](#) [cpg.perm.sort](#) [cpg.perm.sort](#) [cpg.qc](#) [cpg.assoc](#)

Examples

```
data(samplecpg, samplepheno, package="CpGassoc")
results<-cpg.assoc(samplecpg, samplepheno$weight, large.data=FALSE)

cpg.GC(results)
##If the genomic inflation factor is less than one there is no need for adjustment
```

cpg.perm	<i>Perform a Permutation Test of the Association Between Methylation and a Phenotype of Interest</i>
----------	--

Description

Calls `cpg.assoc` to get the observed P-values from the study and then performs a user-specified number of permutations to calculate an empirical p-value. In addition to the same test statistics computed by `cpg.assoc`, `cpg.perm` will compute the permutation p-values for the observed p-value, the number of Holm significant sites, and the number of FDR significant sites.

Usage

```
cpg.perm(beta.values, indep, covariates = NULL, nperm, data = NULL, seed = NULL,
logit.transform = FALSE, chip.id = NULL, subset = NULL, random = FALSE,
fdr.cutoff = 0.05, fdr.method = "BH", large.data=TRUE)
```

Arguments

beta.values	A vector, matrix, or data frame containing the beta values of interest (1 row per CpG site, 1 column per individual).
indep	A vector containing the main variable of interest. <code>cpg.assoc</code> will evaluate the association between <code>indep</code> and the beta values.
covariates	A data frame consisting of the covariates of interest. <code>covariates</code> can also be a matrix if it is a model matrix minus the intercept column. It can also be a vector if there is only one covariate of interest. Can also be a formula (e.g. <code>~cov1+cov2</code>).
nperm	The number of permutations to be performed.
data	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in <code>data</code> , the variables are taken from the environment from which <code>cpg.perm</code> is called.
seed	The required seed for random number generation. If not input, will use R's internal seed.

logit.transform	logical. If TRUE, the logit transform of the beta values $\log(\text{beta.val}/(1-\text{beta.val}))$ will be used. Any values equal to zero or one will be set to the next smallest or next largest value respectively; values <0 or >1 will be set to NA.
chip.id	An optional vector containing chip, batch identities, or other categorical factor of interest to the researcher. If specified, chip id will be included as a factor in the model.
subset	An optional logical vector specifying a subset of observations to be used in the fitting process.
random	logical. If TRUE, the 'chip.id' will be processed as a random effect, and a random intercept model will be fitted.
fdr.cutoff	The threshold at which to compare the FDR values. The default setting is .05. Any FDR values less than .05 will be considered significant.
fdr.method	Character. Method used to calculate False Discovery Rate. Can be any of the methods listed in <code>p.adjust</code> or "qvalue" for John Storey's qvalue method (required to have qvalue package installed). The default method is "BH" for the Benjamini & Hochberg method.
large.data	Logical. Enables analyses of large datasets. When <code>large.data=TRUE</code> , <code>cpg.assoc</code> avoids memory problems by performing the analysis in chunks.

Value

The item returned will be of class "cpg.perm". It will contain all of the values of class `cpg` (`cpg.assoc`) and a few more:

permutation.matrix	A matrix consisting of the minimum observed P-value, the number of Holm significant CpG sites, and the number of FDR significant sites for each permutation.
gc.permutation.matrix	Similar to the <code>permutation.matrix</code> only in relation to the genomic control adjusted p-values.
perm.p.values	A data frame consisting of the permutation P-values, and the number of permutations performed.
perm.tstat	If one hundred or more permutations were performed and <code>indep</code> is a continuous variable, consists of the quantile .025 and .975 of observed t-statistics for each permutation, ordered from smallest to largest. <code>perm.tstat</code> is used by <code>plot.cpg.perm</code> to compute the confidence intervals for the QQ plot of t-statistics. Otherwise NULL.
perm.pval	If one hundred or more permutations were performed, consists of the observed p-values for each permutation, ordered from smallest to largest. <code>perm.pval</code> is used by <code>plot.cpg.perm</code> to compute the confidence intervals for the QQ plot of the p-values. Otherwise NULL.

Author(s)

Barfield, R.; Conneely, K.; Kilaru, V.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.assoc](#) [cpg.work](#) [plot.cpg](#) [scatterplot.cpg](#) [combine](#) [manhattan](#) [plot.cpg.perm](#) [sort.cpg.perm](#)
[sort.cpg](#) [cpg.qc](#)

Examples

```
##Loading the data
data(samplecpg,samplepheno,package="CpGassoc")
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
#Performing a permutation 10 times
Testperm<-cpg.perm(samplecpg[1:200,],samplepheno$weight,seed=2314,nperm=10,large.data=FALSE)
Testperm
#All the contents of CpGassoc are included in the output from Testperm

#summary function works on objects of class cpg.perm
summary(Testperm)
```

cpg.qc

Performs quality control on Illumina data.

Description

cpg.qc is designed to perform quality control on Illumina data prior to analysis. In addition to the matrix of beta values, this function requires as input matrices of Signal A, Signal B, and detection p-values. It will remove samples that have low intensity (mean signal intensity less than half of the overall median or 2000). It can also set to NA datapoints with detection p-values exceeding a user-specified cutoff, and can remove samples or sites that have a missing rate above a user-specified value. Finally, users can opt to compute beta values as $M/(U+M)$ or $M/(U+M+100)$.

Usage

```
cpg.qc(beta.orig,siga,sigb,pval,p.cutoff=.001,cpg.miss=NULL,sample.miss=NULL,
constant100=FALSE,sig.return=FALSE)
```

Arguments

beta.orig	The original beta values matrix obtained from GenomeStudio.
siga	The unmethylated signals matrix obtained from GenomeStudio.
sigb	The methylated signals matrix obtained from GenomeStudio.
pval	A matrix of detection p-values obtained from GenomeStudio. pval should have the same dimension as the beta values and signals: one row for each site and one column for each individual.
p.cutoff	The user-specified cutoff for detection p-values (default=.001).

cpg.miss	Optional cutoff value. If specified, cpg.qc will remove cpg sites where the proportion of missing values exceeds this cutoff.
sample.miss	Optional cutoff value. If specified, cpg.qc will remove samples where the proportion of missing values exceeds this cutoff.
constant100	Logical. If true, the new beta values will be calculated as $M/(U+M+100)$; if false (default) they will be calculated as $M/(U+M)$.
sig.return	Logical. If true, cpg.qc returns the qc'd signal data as well.

Details

It is important that all the matrices or data frames listed above ('pval', 'siga', 'sigb', 'beta.orig') are ordered similarly with respect to samples and CpG sites.

Value

cpg.qc returns a new matrix of beta values that has been subjected to the specified quality control filters. This matrix can be input directly into cpg.assoc.

Author(s)

Barfield, R.; Conneely, K.; Kilaru, V.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.work](#) [cpg.perm](#) [plot.cpg](#) [scatterplot](#) [cpg.combine](#) [cpg.assoc](#)

cpg.work

Does the analysis between the CpG sites and phenotype of interest

Description

Association Analysis Between Methylation Beta Values and Phenotype of Interest. This function contains the code that does the brunt of the work for cpg.assoc and cpg.perm.

Usage

```
cpg.work(beta.values, indep, covariates = NULL, data = NULL, logit.transform = FALSE,
chip.id = NULL, subset = NULL, random = FALSE, fdr.cutoff = 0.05, callarge = FALSE,
fdr.method = "BH", logitperm = FALSE, big.split=FALSE)
```


Arguments

beta.values	A vector, matrix, or data frame containing the beta values of interest (1 row per CpG site, 1 column per individual).
indep	A vector containing the main variable of interest. <code>cpg.work</code> will evaluate the association between <code>indep</code> and the beta values.
covariates	A data frame consisting of the covariates of interest. <code>covariates</code> can also be a matrix if it is a model matrix minus the intercept column. It can also be a vector if there is only one covariate of interest. Can also be a formula (e.g. <code>~cov1+cov2</code>).
data	an optional data frame, list or environment (or object coercible by <code>as.data.frame</code> to a data frame) containing the variables in the model. If not found in <code>data</code> , the variables are taken from the environment from which <code>cpg.work</code> is called.
logit.transform	logical. If TRUE, the logit transform of the beta values $\log(\text{beta.val}/(1-\text{beta.val}))$ will be used. Any values equal to zero or one will be set to the next smallest or next largest value respectively; values <0 or >1 will be set to NA.
chip.id	An optional vector containing chip, batch identities, or other categorical factor of interest to the researcher. If specified, chip id will be included as a factor in the model.
subset	an optional logical vector specifying a subset of observations to be used in the fitting process.
random	logical. If TRUE, the 'chip.id' will be included in the model as a random effect, and a random intercept model will be fitted. If FALSE, 'chip.id' will be included in the model as an ordinary categorical covariate, for a much faster analysis.
fdr.cutoff	The threshold at which to compare the FDR values. The default setting is .05. Any FDR values less than .05 will be considered significant.
callarge	logical. Used by <code>cpg.assoc</code> when it calls <code>cpg.work</code> . If TRUE it means that <code>beta.values</code> is actually split up from a larger data set and that <code>memory.limit</code> may be a problem. This tells <code>cpg.work</code> to perform more <code>rm()</code> and <code>gc()</code> to clear up space.
fdr.method	Character.Method used to calculate False Discovery Rate. Can be any of the methods listed in <code>p.adjust</code> or "qvalue" for John Storey's qvalue method (required to have qvalue package installed). The default method is "BH" for the Benjamini & Hochberg method.
logitperm	Passes from <code>cpg.perm</code> when permutation test is performed. Stops from future checks involving the logistic transformation.
big.split	Passes from <code>cpg.assoc</code> . Internal flag to inform <code>cpg.work</code> that the large data did not need to be split up.

Details

`cpg.work` does the analysis between the methylation and the phenotype of interest. It is called by `cpg.assoc` to do the brunt of the work. It can be called itself with the same input as `cpg.assoc`, it just cannot handle large data sets.

Value

`cpg.work` will return an object of class "cpg". The functions `summary` and `plot` can be called to get a summary of results and to create QQ plots. The output is in the same order as the original input. To sort it by p-value, use the `sort` function.

<code>results</code>	A data frame consisting of the statistics and P-values for each CpG site. Also has the adjusted p-value based on the <code>fdr.method</code> and whether the site was Holm significant.
<code>Holm.sig</code>	A list of sites that met criteria for Holm significance.
<code>FDR.sig</code>	A data.frame of the sites that were FDR significant by the <code>fdr</code> method.
<code>info</code>	A data frame consisting of the minimum P-value observed, the <code>fdr</code> method used, what the phenotype of interest was, and the number of covariates in the model.
<code>indep</code>	The main phenotype of interest.
<code>covariates</code>	If <code>covariates</code> was non NULL, the covariates will be included. Otherwise will be NULL.
<code>chip</code>	If <code>chip.id</code> was non NULL, the chip will be included. Otherwise will be NULL.
<code>coefficients</code>	A data frame consisting of the degrees of freedom, and if object is continuous the intercept effect adjusted for possible covariates in the model, the estimated effect size, and the standard error. The degrees of freedom is used in <code>plot.cpg</code> to compute the genomic inflation factors.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.perm](#) [cpg.assoc](#) [plot.cpg](#) [scatterplot.cpg](#) [combine](#) [manhattan](#) [plot.cpg.perm](#) [sort.cpg.perm](#)
[sort.cpg](#) [cpg.qc](#) [cpg.GC](#)

Examples

```
##See the examples listed in cpg.assoc for ways in which to use cpg.work.
##Just change the cpg.assoc to cpg.work.
```

design

Create full and reduced design matrices for the cpg.assoc function.

Description

Designed to be used by `cpg.assoc` and `cpg.perm`. Creates a full and reduced design matrices.

Usage

```
design(covariates, indep, chip.id, random)
```

Arguments

<code>covariates</code>	A data frame consisting of the covariates of interest. <code>covariates</code> can also be a matrix if it is a model matrix minus the intercept column. It can also be a vector if there is only one covariate of interest. If no covariates must be specified as <code>NULL</code> .
<code>indep</code>	A vector containing the main variable of interest. <code>cpg.assoc</code> will evaluate the association between <code>indep</code> and the beta values.
<code>chip.id</code>	An optional vector containing chip or batch identities. If specified, <code>chip.id</code> will be included as a factor in the model.
<code>random</code>	Is the model going to be a mixed effects. If so, <code>chip.id</code> will not be included in the design matrices.

Value

Returns a list containing the full and reduced design matrices.

<code>full</code>	The full design matrix.
<code>reduced</code>	The reduced design matrix.

Note

The design function is designed to be used exclusively by the `cpg.assoc` and `cpg.perm` functions.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.assoc](#) [cpg.perm](#) [cpg.work.plot](#) [cpg.scatterplot](#) [cpg.combine](#) [manhattan.plot](#) [cpg.perm](#)

Examples

```
data(samplecpg, samplepheno, package="CpGassoc")
#Example where there are covariates:
covar<-data.frame(samplepheno$weight, samplepheno$Distance)
test<-design(covar, samplepheno$SBP, samplepheno$chip, FALSE)
dim(test$full)
dim(test$reduced)
test$reduced[1:5,1:5]
test$full[1:5,1:5]
#When no covariates or chip.id:
test2<-design(NULL, samplepheno$SBP, NULL, FALSE)
dim(test2$full)
dim(test2$reduced)
```

manhattan

*Create a manhattan plot***Description**

This function will produce a manhattan plot for the observed P-values from a object of class "cpg" or "cpg.perm".

Usage

```
manhattan(x, cpname, chr, pos, save.plot = NULL, file.type="pdf",
popup.pdf = FALSE, eps.size = c(15, 5), main.title = NULL, cpg.labels = NULL,
chr.list = NULL, color.list = NULL, point.size = NULL, ...)
```

Arguments

x	Object of class "cpg" or "cpg.perm".
cpname	A vector consisting of the labels for each CpG site.
chr	A vector consisting of the chromosome number for each CpG site.
pos	The map position of each CpG site within its chromosome.
save.plot	Name of the file for the plot to be saved to. If not specified, plot will not be saved.
file.type	Type of file to be saved. Can either be "pdf" or "eps". Selecting file.type="eps" will result in publication quality editable postscript files that can be opened by Adobe Illustrator or Photoshop.
popup.pdf	TRUE or FALSE. If creating a pdf file, this indicates if the plot should appear in a popup window as well. If running in a cluster-like environment, best to leave FALSE.
eps.size	Vector indicating the size of .eps file (if creating one). Corresponds to horizontal and height.
main.title	Main title to be put on the graph. If NULL one based on the analysis will be used.
cpg.labels	A character scalar of either "FDR" or "HOLM" which will label the significant sites on the manhattan plot.
chr.list	A vector listing the chromosomes to be plotted (all available chromosomes are plotted by default). The X and Y chromosomes can be denoted by 23 and 24
color.list	A vector of custom colors to be used for each chromosomes in the manhattan plot.
point.size	The Size of the points in the manhattan plot, if NULL, default to our default, where significant probes have different sizes.
...	Arguments to be passed to methods, such as graphical parameters.

Note

'cpgname', 'chr', and 'pos' must be sorted in the same order, so that the first cpgname[1] corresponds to chr[1] and pos[1], and so on.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.perm](#) [plot.cpg](#) [scatterplot](#) [cpg.assoc](#) [plot.cpg.perm](#) [manhattan.reflect](#)

Examples

```
#Doing a Manhattan plot. First load the data:
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
data(samplecpg, samplepheno, annotation, package="CpGassoc")

examplemanhat<-cpg.assoc(samplecpg, samplepheno$Disease, large.data=FALSE)

manhattan(examplemanhat, annotation$TargetID, annotation$CHR, annotation$MAPINFO)
```

manhattan.reflect *Create a Reflective Manhattan plot*

Description

This function will produce a reflective manhattan plot for the observed P-values from an object of class "cpg" or "cpg.perm". The original analysis needs to be performed on a continuous variable (need T-statistics).

Usage

```
manhattan.reflect(x, cpgname, chr, pos, save.plot = NULL, file.type="pdf",
  popup.pdf = FALSE, eps.size = c(15, 5), main.title = NULL, cpg.labels = NULL,
  chr.list = NULL, color.list = NULL, fdr.cutoff=NULL, point.size=NULL, ...)
```

Arguments

x	Object of class "cpg" or "cpg.perm".
cpgname	A vector consisting of the labels for each CpG site.
chr	A vector consisting of the chromosome number for each CpG site.

pos	The map position of each CpG site within its chromosome.
save.plot	Name of the file for the plot to be saved to. If not specified, plot will not be saved.
file.type	Type of file to be saved. Can either be "pdf" or "eps". Selecting file.type="eps" will result in publication quality editable postscript files that can be opened by Adobe Illustrator or Photoshop.
popup.pdf	TRUE or FALSE. If creating a pdf file, this indicates if the plot should appear in a popup window as well. If running in a cluster-like environment, best to leave FALSE.
eps.size	Vector indicating the size of .eps file (if creating one). Corresponds to horizontal and height.
main.title	Main title to be put on the graph. If NULL one based on the analysis will be used.
cpg.labels	A character scalar of either "FDR" or "HOLM" which will label the significant sites on the manhattan plot.
chr.list	A vector listing the chromosomes to be plotted (all available chromosomes are plotted by default). The X and Y chromosomes can be denoted by 23 and 24
color.list	A vector of custom colors to be used for each chromosomes in the manhattan plot.
fdr.cutoff	A numeric scalar between 0 and 1 to indicate what to consider FDR significant. Defaults to NULL.
point.size	The Size of the points in the manhattan plot, if NULL, default to our default, where significant probes have different sizes.
...	Arguments to be passed to methods, such as graphical parameters.

Note

'cpgname', 'chr', and 'pos' must be sorted in the same order, so that the first cpgname[1] corresponds to chr[1] and pos[1], and so on.

Author(s)

Barfield, R.; Kilaru, V.; Conneely, K.
 Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.perm plot](#) [cpg.scatterplot](#) [cpg.assoc plot](#) [cpg.perm manhattan](#)

Examples

```
#Doing a Manhattan plot. First load the data:
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
data(samplecpg, samplepheno, annotation, package="CpGassoc")

examplemanhat<-cpg.assoc(samplecpg, samplepheno$Disease, large.data=FALSE)
```

```
manhattan.reflect(examplemanhat, annotation$TargetID, annotation$CHR, annotation$MAPINFO)
```

Other CpGassoc Functions

Information on miscellaneous other functions

Description

`cpg.length` compares the dimensions of the covariates, the independent phenotype, chip, and the matrix of beta values from `cpg.assoc` or `cpg.perm`. If the number of individuals does not match up. Stops the code.

`pointsizefunction` simply scales the size of the points for the qq-plot for `plot.cpg` and `plot.cpg.perm`

`cpgassocsummary` is used by `cpg.work` when the phenotypes contains such small amount of variance such that matrix methods can not be used. Takes an object of class "aov" or "mlm" and gets the test statistics from these objects while avoiding doing a loop or the list structures which is typically done if one does summary on one of these objects.

Usage

```
cpg.length(indep, numpatients, covariates, chip.id)
```

```
pointsizefunction(x)
```

```
cpgassocsummary(object)
```

Arguments

<code>indep</code>	object indep from <code>cpg.assoc</code> , <code>cpg.work</code> or <code>cpg.perm</code> .
<code>numpatients</code>	The number of column of the object <code>beta.values</code> in <code>cpg.work</code> or <code>cpg.perm</code> or number of columns of <code>beta.val</code> in <code>cpg.assoc</code>
<code>covariates</code>	object covariate from <code>cpg.assoc</code> , <code>cpg.work</code> or <code>cpg.perm</code> .
<code>chip.id</code>	object <code>chip.id</code> from <code>cpg.assoc</code> , <code>cpg.work</code> or <code>cpg.perm</code> .
<code>x</code>	the p-values from the object "cpg" or "cpg.perm"
<code>object</code>	An object of "aov" or "mlm"

Details

`cpg.length` stops the functions if the dimensions do not add up. Used Internally by **CpGassoc**

`pointsizefunction` simply returns a vector of equal length of `x` to be used in plot for the `cex` option.

`cpgassocsummary` returns a matrix of columns two and three of the results value of an "cpg" or "cpg.perm" object. This is for the cpg sites with non-missing data

See Also

[cpg.perm](#) [cpg.assoc](#) [scatterplot](#) [manhattan](#) [plot.cpg.perm](#)

Sample Data CpGassoc *Sample data from CpGassoc*

Description

- samplecpgMatrix containing sample (fake) methylation data for 258 individuals over 1228 CpG sites
- samplephenoMatrix with phenotype info for the 258 individuals
- annotationMatrix with annotation information for the 1228 CpG sites

Usage

```
data(samplecpg)
```

Format

- samplecpgMatrix, dimensions 1228 x 258
- samplephenoMatrix, dimensions 258 x 6. Phenotype info on "Dose", "SBP", "Distance", "weight", "Disease", and "chip"
- annotationMatrix, dimensions 1228 x 3. Header names are "TargetID", "CHR", and "MAPINFO"

See Also

[cpg.perm](#) [cpg.assoc](#) [scatterplot](#) [manhattan](#) [plot.cpg.perm](#)

Examples

```
##See help pages for other functions for usage of these datasets
```

scatterplot *Plot beta values of individual CpG sites against the independent variable.*

Description

Plot beta values of individual CpG sites against the independent variable. Can create scatterplots and boxplots. If scatterplots the intercept will be adjusted for any covariates that were included in the model.

Usage

```
scatterplot(x, cpg.rank = NULL, cpg.name = NULL, save.plot = NULL, file.type="pdf",
eps.size = c(5, 5), popup.pdf = FALSE, beta.values = NULL,main.title=NULL, ...)
```

Arguments

x	Object of class "cpg" or "cpg.perm".
cpg.rank	A vector listing the rank of sites to be plotted. The rank is based on the ordered p-values.
cpg.name	A character vector containing the names of CpG sites to be plotted against the phenotype of interest. This option is ignored if 'cpg.rank' is specified.
save.plot	Prefix of the filename for the plot(s) to be saved to. If specified, plot filenames will be created by appending this prefix to either cpg.rank or cpg.name. If not specified, plot will not be saved.
file.type	Type of file to be saved. Can either be "pdf" or "eps". Selecting file.type="eps" will result in publication quality editable postscript files that can be opened by Adobe Illustrator or Photoshop.
eps.size	Vector indicating the size of .eps file (if creating one). Corresponds to horizontal and height.
popup.pdf	TRUE or FALSE. If creating a pdf file, this indicates if the plot should appear in a popup window as well. If running in a cluster-like environment, best to leave FALSE.
beta.values	If the object has been renamed (i.e. x\$info\$beta\$info is no longer in ls(.GlobalEnv)) then specify the new object here.
main.title	Main title to be put on the graph. If NULL one based on the analysis will be used
...	Arguments to be passed to methods, such as graphical parameters.

Details

An unlimited number of CpG sites can be selected for plotting by specifying either 'cpg.rank' or 'cpg.name', as shown in the Examples below. Note that only one of these options is needed; if both are entered, 'cpg.rank' will be used.

Author(s)

Barfield, R.; Kilaru,V.; Conneely, K.
Maintainer: R. Barfield: <rbarfield01@fas.harvard.edu>

See Also

[cpg.assoc.plot.cpg.manhattan](#) [cpg.perm.plot.cpg.perm](#)

Examples

```
#Load the data:
data(samplecpg,samplepheno,package="CpGassoc")
###NOTE: If you are dealing with large data, do not specify large.data=FALSE.
###The default option is true.
##This will involve partitioning up the data and performing more gc() to clear up space
test<-cpg.assoc(samplecpg,samplepheno$weight,large.data=FALSE)
##Using rank, will plot the top three sites in order of significance:
scatterplot(test,c(1:3))
##Using name, specify three sites:
scatterplot(test,cpg.name=c("CpG1182","CpG1000","CpG42"))

##Plotting something that is categorical in nature:
test2<-cpg.assoc(samplecpg[1:200,],factor(samplepheno$Disease),large.data=FALSE)
scatterplot(test2,c(2),beta.values=samplecpg[1:200,])
```

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